١	Integron/gene cassette metagenome in marine environments, exploring and applications.
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1 5	A review article
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Abstract

۲ Recently, integron gene cassette metagenome has been discovered as a class of mobile ٣ DNA elements that play a role in the functional genome of marine bacteria. This importance of ٤ integron was extended from the original idea that integron is the main key for evolution and ٥ adaptation of pathogenic bacteria against antibiotic attacks. In this review, we highlighted on the ٦ diversity, function and applications of integron/gene cassette metagenome in marine ٧ environments. Each environment has specific selective characteristics that could shape the ٨ integron gene cassette metagenome, to be an environment adaptive DNA tool. This idea led us to ٩ innovate synthetic integrons with wide biotechnological applications.

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1. Introduction

11 The genomics era has clearly indicated that a large proportion of marine bacterial genes have been acquired by horizontal gene transfer [Ochman et al., Y...]. This type of gene transfer ۱۲ ۱۳ is facilitated by a number of genetic elements, including plasmids, transposons, and integrons. Traditionally, most attention has focused on plasmids and transposons [Smalla et al., ⁷...]. ١٤ 10 However, since integrons have recently been demonstrated to occur in the genome of diverse ١٦ marine uncultured bacterial species and integron integrases are recoverable from various marine environmental samples [Elsaied et al., ^Y··^Y; Rodríguez-Minguela et al., ^Y··⁹]; we reasoned that ۱۷ integrons are widespread in marine environments. ۱۸

Integron is DNA element described in pathogenic and environmental bacteria, first
 recognized and named by Ruth Hall and Hatch Stokes in 1949 [Stokes and Hall, 1949]. Integron
 carries varieties of functional expressed genes, in the form of cassettes, which play an important
 process in shaping bacterial genomes over evolutionary time scales. In particular, in the context
 of the short-term evolution of bacterial genomes, mechanisms of non-homologous recombination

١ are known to be crucial to adaptation in extreme environments. The most obvious example of ۲ this is the extraordinary ability of pathogens to acquire multiple antibiotic-resistance genes in ٣ historical times, a feature characterizing only integron metagenome. Recently, integron was ٤ found to have the flexibility to acquire and expel genetic elements adaptive to several stresses, ٥ other than antibiotics, in marine environment [Elsaied et al., $\forall \cdot \cdot \forall$; Koenig et al., $\forall \cdot \cdot \uparrow$]. This ٦ integron unique feature created a system that provides for an enormous pool of known and ۷ unknown adaptive genes to be mobilized, rearranged, and disseminated amongst marine bacteria. Indeed, the reservoir of adaptive genes capable of being mobilized as yet has no known upper ٨ ٩ limit but must, at the very least, number in the thousands and is probably orders of high ۱. magnitude. As a result, integron system can greatly influence marine bacterial diversity and ۱١ adaptation in ways that other DNA tools cannot.

١٢ In order to understand the integron metagenome contribution to the evolution and ۱۳ adaptation of marine bacteria, we looked at extreme marine environments where the bacterium ١٤ exposes to various selective environmental pressures, which stimulate integron diversity and 10 abundance. We described the diversity and shape of integron metagenome in two types of ١٦ extreme marine environments. The first is deep-sea hydrothermal vents, environments ۱۷ completely away from anthropogenic activities, where both free-living and symbiotic bacteria ۱۸ expose to different selective natural pressures such as high temperature and toxic gases erupted ۱٩ from the vents [Cherry et al., 1997]. The second constitutes the urban marine sediments, which ۲. are exposing to a high load of different human industrial waste pollutions.

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Y. Integron structure in marine environment

Integron is a site-specific recombination system. The ideal integron consists of three
 DNA structures, which are integrase gene, *intI*, attachment site, *attI*, and integrated gene

cassettes (Fig. 1). The integrase gene, *intI*, encodes the enzyme integrase, belonging to the
 tyrosine recombinase family, which is responsible for integration of gene cassettes inside
 integron [Esposito and Scocca, 1997].

٤ Like other enzymes, integrase, IntI, has specific motive catalytic sequence, which encodes the conserved amino acid residues [RHRY] found in two sequence boxes [I and II] ٥ ٦ located in the carboxylic half of the integrase protein [Nancy and Roy Y...)]. *intI* was identified ۷ in marine integron metagenomes through PCR amplification and sequencing of only fragments from the *intI* terminal part, which encodes the carboxylic half of the expressed enzyme. ٨ ٩ Molecular culture-independent studies obtained *intI* sequences of size $\forall 1 \notin bp$ ($\forall \cdot \%$ of full ideal *intI* sequence $\sim 1 \cdot 7 \cdot \text{ bp}$) from varieties of free-living and symbiotic hydrothermal vent bacterial ۱. ۱۱ metagenomes [Elsaied et al., $\gamma \cdot \gamma$, $\gamma \cdot \gamma$], while only $\xi \gamma \gamma$ bp of *intl* were sequenced from ۱۲ urban shore and Antarctic marine sediment metagenomes [Minguela-Rodrigues et al., Y., 9]. *intl* ۱۳ sequences recovered from marine metagenome contained some of the catalytic conserved intI ١٤ regions founded in clinical and cultured isolates [Grainge and Jayaram, 1999], implicating the ۱٥ global consensus motive structure of *intI*.

The promoter, called P_c , located upstream to *intI*, [Collis and Hall, 1990] and responsible for expression of integrated gene cassettes, has yet to be characterized experimentally in marine integron metagenome.

The attachment site, attI, is the integron-associated recombination site, where the gene cassettes are captured by integrons [Collis et al., $\forall \cdot \cdot \forall$] (Fig. 1). Thirty three diverse attI-like sequences have been detected in marine integrons, and most contained an integrase-binding simple site, but lacked a pair of direct repeat sequences as found in attI of known integrons in pathogenic bacteria (Partridge et al., $\forall \cdot \cdot \cdot$; Elsaied et al., $\forall \cdot 1 \rangle$, $\forall \cdot 1 \forall$).

١ The units of insertion into integrons are mobile gene cassettes. A gene cassette is an ۲ independently mobilizable element that generally contains a promoterless ORF and an intI-٣ recognizable recombination site called the *attC* [Stokes et al., $\gamma \cdots \gamma$]. In several integrated gene ٤ cassettes recorded from marine metagenomes, *attC* has been considered as part of the ORF, while in others it is located downstream to the ORF [Elsaied et al., $\gamma \cdot \cdot \gamma$; Koenig et al., $\gamma \cdot \cdot \lambda$, ٥ ٦ $\gamma \cdot \cdot \gamma$]. The *attC* sites have diverse sequences and lengths in both clinical and environmental ۷ isolates but share common features including about $\gamma \circ$ bp at each end that conform to consensus sequences [Nield et al., $\gamma \cdots \gamma$; Stokes et al., $\gamma \cdots \gamma$]. The *attC* consensus regions are ٨ ٩ imperfect inverted repeats of one another, and each comprises a pair of inversely oriented ۱. integrase-binding domains, separated by a spacer of \vee or \wedge bp [Hall et al., \vee \vee] (Fig. \vee). The ۱۱ recombination crossover has been localized between the G and first T of a seven-base core site ۱۲ (region R in **Fig.**) located in the right-hand simple site. Recombination activity of *attC* from ۱۳ deep-sea integron metagenome has been measured experimentally (Elsaied et al., $\forall \cdot \cdot \forall$). The ١٤ lengths of *attC* recorded from marine metagenomes were ranged from \circ^{9} bp to $\gamma \gamma$, bp, an implication for enormous diversity of *attC* in marine environment [Elsaied et al., Y.)), Y.)"; 10 Koenig et al., $\gamma \cdot \cdot \wedge$, $\gamma \cdot \cdot \circ$; Wright et al., $\gamma \cdot \cdot \wedge$]. ١٦

There is no obvious evidence for the origin of marine integron metagenome. However, integron is an old genetic element in the evolution of bacterial genome and not related, in origin, to the short history of antibiotics. This concept is confirmed by the discovery of integron in deepsea bacteria, representing the oldest form of bacterial genome in our biosphere and completely isolated from any effect of antibiotics [Elsaied et al., $\forall \cdot \cdot \forall$]. On the other hand, lateral gene transfer through bacterial conjugation may help in the evolutionary spreading of integrons

between clinical pathogenic and marine bacteria through disposable wastes of human into
marine environment [Wright et al., ۲۰۰۸; Koenig et al., ۲۰۰۹].

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*****. Diversity of integrase gene, *intI*, as a marker for integron diversity in marine environments

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Two features supported studying the diversity of integron based on integrase gene. The
 first feature is the common existence of integrase gene as the backbone of any integron
 recovered from both of clinical and environmental bacteria. The second feature is conservation
 of the integrase motive sequence in almost all recovered bacterial integrons [Nancy and Roy,
 Y···Y].

۱۱ Originally, three major groups (class 1 to 7) of integron integrase are highly prevalent in ۱۲ the clinical scene [Mazel et al., 199A]. In most of the cases, these three classes of integrase have ۱۳ also been reported to catch up almost exclusively gene cassettes encoding antibiotic resistance ١٤ functions [Mazel, $\forall \cdot \cdot \forall$]. All together, these traits have led to their designation as mobile clinical 10 integron integrases [Gillings et al., $\forall \cdot \cdot \wedge$]. Although integrons have been traditionally classified ١٦ according to the percent identity of the nucleotide or predicted amino acid sequence of their ١٧ respective integrase genes, several structural features and differences in abundance patterns have been identified and distinguished classes 1 to 7 [Biskri et al., $7 \cdot \cdot \circ$; Xu et al., $7 \cdot \cdot \sqrt{1}$]. ۱۸

Class ' integron integrase, *intI*', is the most widely studied variant and is typically linked to replicative Tn^{r} transposons, which appear to contribute to their extensive distribution [Nandi et al., $^{r} \cdot \cdot ^{\epsilon}$]. The diversity of class ' integron has been extended beyond clinical field. The relative *intI*' abundance, based on qPCR, was high in bacterial metagenomes isolated from polluted estuary and shore sediments [Gillings et al., $^{r} \cdot \cdot ^{k}$; Wright et al., $^{r} \cdot \cdot ^{k}$], implicating the role of lateral gene transfer in spreading of this class in both clinical and marine bacteria. Recently, we discovered class ' integrase gene in a deep-sea symbiont living within a gutless
 clam, *Calyptogena* sp. (accession no. AB^{YYYYYY}) [Elsaied et al., unpublished data], Sagami bay,
 Japan, and recording amino acid identity ^{AY}? with that of *Escherichia coli* (Fig. "), a feature
 supporting the concept that this class has an ancient phylogenetic lineage in bacterial genome.

٥ In contrast to class intl¹, class ^Y integrase, intl^Y, is routinely associated with non-٦ replicative Tn^{V} transposons [Ahmed et al., $\uparrow \cdot \cdot \uparrow$; Barlow and Gobius, $\uparrow \cdot \cdot \uparrow$; Marquez et al., ۷ $\{\cdot,\cdot\}$. Class $\{\cdot,\cdot\}$. Class $\{\cdot,\cdot\}$ have been found to be abundant in sediments with high input of sewage and fecal wastes, while poorly represented in the environments with moderate or no ٨ ٩ recent anthropogenic impact [Minguela-Rodriguez et al., $\gamma \cdot \cdot \gamma$]. The presence of *intl* γ -like ۱. integrase in environments impacted by fecal waste from animals, including humans, allows us to ۱۱ suggest that *intI* γ may have been extracted from the extant environmental pool, presumably ۱۲ through the food chain and in the absence of antibiotic selection, and then laterally transferred ۱۳ and enriched in the gastrointestinal tract. These events seem feasible, as the ability of exogenous ١٤ bacteria to survive transit in the human and animal gut environment and the occurrence of 10 bacterial lateral gene transfer in the marine setting are well documented [Shoemaker et al., ^Y··^Y; Minguela-Rodriguez et al., $\gamma \cdot \cdot \gamma$]. Even less is known about the class γ variants, which so far ١٦ have been described in only three clinical instances [Xu et al., $\forall \cdot \cdot \forall$]. ۱۷

Recent studies on marine integron metagenomes recorded high diverse integrases that have unique phylogenetic lineages (**Fig.** (*)), expanding the diversity of *intI* beyond the three traditional classes. More than (*) *intI* phylotypes were recorded in metagenomes isolated from varieties of marine environments including hydrothermal vent fluids, symbionts and marine sediments from Arctic [Elsaied et al., (*, *); Minguela-Rodriguez et al., (*, *)], a fact concerning the wide biogeographic distribution of integron in marine environments. The most interesting feature of these *intI* diversities is the phylogenetic distinction of *intI* phylotypes between the
 studied environments. This may refer to the wide geographic distance and topology between the
 studied marine habitats.

Despite the large diversity of the recorded *intI*, the expectation of real diversity of *intI* in
the studied marine environments has no upper limit as most of the studied *intI* clone libraries
have yet to be saturated. More *intI* monitoring and improving techniques like developing new
efficient *intI* PCR primers and qPCR will expand our knowledge about diversity of *intI* in marine
environments.

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4. How big is the gene cassette pool in marine environment?

Culture-independent PCR/sequencing approaches yielded more than $\tau \cdots$ gene ۱. cassettes from different marine integron metagenomes including Y120, Y.A, 17V and 27 ۱۱ cassettes from oil and sewage polluted marine sediments, coal and steel contaminated ۱۲ ۱۳ shores, deep-sea and heavy metals polluted estuaries, respectively, (Fig. [£]) [Elsaied et al., $\gamma \cdot \gamma$, $\gamma \cdot \gamma$; Koenig et al., $\gamma \cdot \gamma$; Wright et al., $\gamma \cdot \gamma$]. However, these ١٤ numbers of gene cassettes represent a tiny fraction of real gene cassette pool in each 10 ١٦ metagenome. This expectation supported by rarefaction analyses, based on gene cassette ۱۷ clone libraries, which indicated that sampling has yet to capture the diversity found in clone libraries from these sampling sites [Elsaied et al., Y.), Y.), Koenig et al., Y. A, ۱۸ $\gamma \cdot \cdot \gamma$: Wright et al., $\gamma \cdot \cdot \lambda$]. It should be considered that cassette analyses were done from ۱٩ ۲. relatively few samples, making these studies a substantial underestimate of marine ۲١ integron gene cassette metagenome diversity. Primers used in these studies were designed from nucleotide alignments of a relatively small subset of *attC* sites [Stokes et al., $\gamma \cdots \gamma$]. ۲۲ However, the recorded *attC* sites are highly variable, demonstrating that the level of ۲۳ ۲٤ diversity of these *attC* elements continues to grow with cassette richness. Moreover, in all

studies done on marine integron gene cassette metagenomes, the used PCR approach was not sufficient to pick the full length of integrated gene cassettes, where the sizes of PCR ampilicons were short, consisted of, mostly, a single gene cassette [Elsaied et al., $\gamma \cdot \gamma$], $\gamma \cdot \gamma \gamma$; Koenig et al., $\gamma \cdot \gamma \gamma$; Wright et al., $\gamma \cdot \gamma$].

Most of the recorded gene cassettes contained ORFs oriented in forward
 expression direction. However, some gene cassettes, with no apparent ORFs and
 contained only *attC*, were recorded in marine integrons. It is certain that more exhaustive
 sampling and additional methodology, in terms of gene cassette data collection, will
 reveal more cassette diversity in the marine environments.

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•. Marine integron gene cassette metagenome is a big reservoir of known and unknown gene cassette-coding proteins

The integrated gene cassettes recorded in marine integron metagenomes have structural features such as shortage of their containing ORFs (from \mathfrak{so} bp to \mathfrak{lo} kb), mostly having ribosomal binding site upstream to the ORF and ORF initial codons ATG, GTG or TTG [Elsaied et al., \mathfrak{rov} ; Koenig et al., \mathfrak{rov} ; Wright et al., \mathfrak{rov}]. These features facilitate easy expression of cassette ORFs, using integron cassette promoter, *Pc*, into proteins, which involve in different bacterial activities.

Most of gene cassette ORFs from clinical field encode antibiotic resistance proteins. Given that the lateral gene transfer plays an efficient role in moving the integron metagenome from clinical to urban marine environments, antibiotic resistance genes from clinical isolates have yet to be recorded in marine sediments. This is may be due to the limitation of gene cassette survey in marine environment. Other explanation is that integron may use its unique feature to disseminate antibiotic resistance genes, when the bacterium moves from clinic to marine environment, where there is no stress of antibiotics. However, more studies are needed to
 confirm this concept.

٣ Blast homology search showed that gene cassette ORFs, picked from marine integron ٤ metagenomes, encoded several categories of proteins (Fig. •). The first category included ٥ environment-adaptive proteins, which may be expressed by integron under exposure of ٦ bacterium to surrounding environmental selective pressures. Xenobiotic-degrading enzymes and ۷ pollutant signal proteins are the most recorded marine environment adaptive cassette proteins (Koenig et al., $\gamma \cdot \gamma$; Elsaied et al., $\gamma \cdot \gamma$). Protein bioinformatics predicted cell membrane ٨ ٩ localization of the cassette signal proteins, an implication for their functions as biosensors to ۱. surrounding environmental stresses. Some cassette adaptive proteins were expressed as a ۱۱ response of specific environment selective pressures, such as heat shock chaperon cassette ۱۲ proteins that may play a role in protection of bacteria from high temperature of deep-sea hydrothermal vents [Elsaied et al., $\forall \cdot \cdot \forall$]. Several gene cassettes encoded catechol \uparrow, ξ_{-} ۱۳ ١٤ dioxygenase (EC 1.17.11.1) and ξ -hydroxy-1-oxovalerate aldolase (EC $\xi.1.7.79$), enzymes 10 involve in degradation of aromatic industrial wastes such as benzoate, toluene and xylene, were ١٦ recorded in coal and steel industrial waste contaminated marine sediment, an implication for ۱۷ involving of cassette proteins in the field of bioremediation [Koenig et al., Y., 9].

Other category consisted of cassette metabolism-related proteins. The most obvious examples of these proteins were transferase enzymes and potassium and iron ion transporter proteins, which were recorded in almost all studied inetgron metagenomes, suggesting the wide biogeographic distribution of these mobile cassette proteins in marine environments [Elsaied et al., $7 \cdot r$, $7 \cdot r$, $7 \cdot r$; Koenig et al., $7 \cdot r$, $7 \cdot r$; Wright et al., $7 \cdot r$]. In addition, deep-sea integron metagenomes were found to be rich with cassette ORFs encoding specific enzymes such

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as glycoside hydrolase, malate dehydrogenase and thioredoxin reductase beside DNA-binding
 enzymes such as transposases, suggesting the role of transposon as integron metagenome carrier
 in deep marine environment [Elsaied et al., Y···V].

Cassette-encoding structural proteins constituted other category in marine gene cassette
 metagenomic pool. Some cassettes encoded novel protein domains and families such as a new
 family of nuclease-related domain (NERD) proteins, which were found in a broad range of
 bacterial, as well as single archaeal and plant proteins. The presence of NERD in the virulence related pXO¹ plasmid of *Bacillus anthracis* as well as in several other pathogens makes it a
 possible drug target [Grynberg and Godzik, ^Y · · ^x; Elsaied et al., ^Y · · ^y].

We have found examples of the IS ξ family of transposons linked to integrons from deep biosphere [Elsaied et al., $\forall \cdot \cdot \forall, \forall \cdot 1 \forall, \forall \cdot 1 \forall$]. Because the deep-sea microbial ecosystem has been considered as a primitive fraction of the biosphere, the transposon, IS ξ family, might be responsible for horizontal transfer of integrons and associated cassette genes in early history. However, this hypothesis would require further study.

The last category included cassette proteins, which have unknown functions and constituted the major fraction of cassette proteins recorded in marine environments. These unknown cassette proteins need characterization based on both protein prediction bioinformatics and experimental analyses. Thus, marine integron gene cassette metagenome has been considered as a source of unlimited number of mobile genes that encode novel proteins, which may have importance in the terms of scientific and commercial values.

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5. Integron metagenome as an active player in deep marine bacterial adaptation, the case is a deep-sea symbiont

The example of utilizing adaptive gene cassette metagenome in deep marine environment
 was represented by a deep-sea hydrothermal vent mussel sulfur-oxidizing endosymbiont. This
 endosymbiont lives inside the gill cells of the host mussel [Cavanaugh, \٩^\"] (Fig. \a). Hence,
 this symbiont is continuously exposing to various surrounding vent selective pressures. The
 endosymbiont utilizes hydrogen sulfide, erupted from the hot vent, as an electron donor for
 assimilation of organic carbon through the process of chemosynthesis [Felbeck et al., \٩^\].

٩ Three groups of integron gene cassettes have been studied in this symbiont and encoded proteins with functions adapted to the symbiont life within the host animal [Elsaied et al., $\forall \cdot \cdot \forall$]. ۱۰ 11 Expression studies were performed on these gene cassettes in order to confirm their protein ۱۲ products and functions (Fig. [\]b) [Elsaied et al., unpublished data]. The first group was found to ۱۳ encode methionine aminopeptidase. Methionine is a sulfur-containing essential amino acid and is used as a source of sulfur in the animal symbiont-containing tissue [Duplessis et al., $7 \cdot \cdot \xi$]. ١٤ 10 Specifically, methionine aminopeptidase helps in liberating sulfur, from methionine, which is ١٦ utilized as an electron sink, instead of oxygen, to produce sulfide during anoxic condition ۱۷ [Duplessis et al., $\forall \cdot \cdot \xi$]. Sulfide production in gut flora from the large intestine in humans has ۱۸ been attributed to this mechanism [Magee et al., ^Y···]. Other gene cassette proteins matched ۱٩ with aminopeptidase N and O-sialoglycoprotein endopeptidase, enzymes playing a role in the ۲. fermentation of amino acids, which have been used, under certain conditions, as a source of ۲١ carbon and energy in several deep-sea hydrothermal vent bacteria [Hou et al., $7 \cdot \cdot \xi$].

The second group of symbiont gene cassettes encoded glutamate synthase. This enzyme
 is a complex iron-sulfur flavoprotein that catalyses the reductive transfer of the amido nitrogen
 from L-glutamine to ⁷-oxoglutarate to form two molecules of L-glutamate, a reaction in the

symbiont metabolic pathway for ammonia assimilation [Lee and Childress, 1995; Minic and
 Herve, Y...5].

٣ The third group included gene cassette proteins matched with DNA repair proteins. ٤ Genomes are subjected to damage by physical and toxic chemical agents in the deep-sea ٥ hydrothermal vent environment. For example, increased temperature at vent sites is known to ٦ have the potential to cause DNA damage [Pruski and Dixon, $\forall \cdot \cdot \forall$]. Another possible cause of ۷ DNA damage is the radioactive materials, which always occur at high levels in hydrothermal ٨ habitats [Cherry et al., 1997]. As the endosymbiont is located in the external gill organ of the ٩ mussel, it is always directly exposed to these harmful conditions. Hence, DNA repair proteins ۱. have been recorded in vent mussel symbionts [Pruski and Dixon, $\forall \cdot \cdot \forall$]. Generally, the presence ۱۱ of these mobile gene cassettes suggested that their associated proteins may be adaptive and used ۱۲ as a community resource in this specialized symbiont environment.

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V. Problems and prospective

Only, very few studies concerning integron gene cassette metagenome in marine environments have been reported. There is still lack information about the complete structure of marine integron metagenome. All previously used PCR primers pick only partial sequences of integrase genes or maximum three gene cassettes from long expected gene cassette arrays. In addition to these methodological limitations, most of the recorded gene cassettes encoded proteins with no significant homology with those in databases and new methodologies must be developed to uncover the functions of these cassette proteins.

However, this review gave background about the discovery of integron metagenome in
 marine environment and highlighted the importance of this mobile metagenome in molecular
 adaptation of marine bacteria. Future studies will focus on two aspects. The first aspect should
 include improvement the monitoring of integron metagenome in wide range of environments,
 supported with new concepts and methodologies. This will help in understanding the role of this
 mobile metagenome in environmental microbial ecosystem dynamics and function.

The second aspect will benefit from the function of integron, in collecting of environment
 adaptive functional genes, in developing synthetic integron recombinant system. This innovative
 system will help in screening adaptive genes useful for both of medical and industrial
 applications.

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References

Ahmed A, Kawaguchi F, Shimamoto T (۲۰۰٦) Class Y integrons in Vibrio cholerae. J. Med.
 Microbiol. oo: ٦٤٣–٦٤٤.

¹⁰ Barlow R, Gobius K ((\cdot, \cdot, \cdot)) Diverse class \cdot integrons in bacteria from beef cattle sources. J.

Antimicrob. Chemother. $\circ A: 1177-117A$.

W Biskri L, Bouvier A, Gue'rout M, Boisnard S, Mazel D ((...) Comparative study of class

integron and *Vibrio cholerae* superintegron integrase activities. J. Bacteriol. 149:195.-190.

Cavanaugh C (1947) Symbiotic chemoautotrophic bacteria in marine invertebrates from sulfide-

 γ rich habitats. Nature $\gamma \cdot \gamma$: $\circ \Lambda$.

Cherry R, Desbruyeres D, Heyraud N (1997) High levels of radioactivity in hydrothermal vent

polychaetes. C R Acad. Sci. Paris Ser III ⁽¹⁾: ⁽¹⁾.

Collis C, Hall R (۱۹۹۵) Expression of antibiotic resistance genes in the integrated cassettes of
 integrons. Antimicrob Agents Chemother. ^۳9: ۱۹۹–۱۹۲.

Collis CM, Kim MJ, Stokes HW, Hall RM (^Y··^Y) Integron encoded IntI integrases preferentially
recognize the adjacent cognate attI site in recombination with a °⁹-be site. Mol Microbiol [£]¹:
) £10-1£TY.

٦

Duplessis M, Ziebis W, Gros O, Caro A, Robidart J, Felbeck H (^(*, *)) Respiration strategies
 utilized by the gill endosymbiont from the host Lucinid *Codakia orbicularis* [Bivalvia:
 Lucinidae]. Appl. Environ. Microbiol. ^(*) : ^(*) : ^(*) : ^(*) : ^(*)

۱۰ Elsaied H, Stokes H, Nakamura T, Kitamura K, Fuse H, Maruyama A (۲۰۰۷) Novel and diverse

integron integrase genes and integron-like gene cassettes are prevalent in deep-sea hydrothermal
 vents. Environmental Microbiology 9: YY9A_YYYY.

IT Elsaied H, Stokes HW, Kitamura K, Kurusu Y, Kamagata Y, Maruyama A (⁽¹⁾) Marine

integrons containing novel integrase genes, attachment sites, attI, and associated gene cassettes

in polluted sediments from Suez and Tokyo Bays. ISME J o: 117-117.

Elsaied H, Stokes H, Yoshioka H, Mitani Y, and Maruyama A ((*)). Novel integrons and gene

v cassettes from a Cascadian submarine gas hydrate-bearing core. FEMS Microbiology Ecology,

In press doi: 1.1111/10V£_19£1.1777V. [Epub ahead of print].

۱۹

*• Esposito D, Scocca J (199V) The integrase family of tyrosine recombinases: evolution of a conserved active site domain. Nucleic Acids Res. Yo: $T1.0-T11\xi$.

۲۲

Felbeck H, Childress J, Somero G (1911) Calvin Benson cycle and sulfide oxidation enzymes in animals from sulfied-rich habitats. Nature 197: 11-197.

١	Gillings M, Krishnan S, Worden P, Hardwick S ((\cdot, \cdot)) Recovery of diverse genes for class \cdot
۲	integron-integrases from environmental DNA samples. FEMS Microbiol. Lett. ۲۸۷: ٥٦–٦٢.
٣	Grainge I, Jayaram M (1999) The integrase family of recombinase: organization and function of
٤	the active site. Mol. Microbiol. $\text{TT}: \xi \xi q = \xi \circ \tau$.
0	Grynberg M, Godzik A, $({}^{\boldsymbol{\tau}} \cdot {}^{\boldsymbol{\xi}})$ NERD: a DNA processing-related domain present in the anthrax
٦	virulence plasmid, pXO ¹ . TRENDS in Biochemical Sciences ^{Y9} : 1.1-11.
٧	Hall R, Brookes D, Stokes H, (1991) Site-specific insertion of genes into integrons: role of the
٨	o9-base element and determination of the recombination cross-over point. Mol. Microbiol.
٩	0:1921_1909.
۱.	Hou S, Saw J, Lee K, Freitas T, Belisle C, Kawarabayasi Y, et al. $(\uparrow \cdot \cdot \not)$ Genome sequence of
• •	the deep-sea g-Proteobacterium idiomarina loihiensis reveals amino acid fermentation as a
۱۲	source of carbon and energy. Proc. Natl. Acad. Sci. USA 1.1: 14.77-14.51.
۱۳	Koenig J, Sharp C, Dlutek M, Curtis B, Joss M, Boucher Y, Doolittle W (^(,,)) Integron gene

cassettes and degradation of compounds associated with industrial waste: the case of the Sydney
 Tar Ponds. PLoS ONE \$, online

Koenig J, Boucher Y, Charlebois R, Nesbø C, Zhaxybayeva O, Bapteste E, Spencer M, Joss

M, Stokes H, Doolittle W $(\uparrow \cdot \cdot \land)$ Integron-associated gene cassettes in Halifax Harbour: assessment of a mobile gene pool in marine sediments. Environmental Microbiology $\uparrow \cdot : \uparrow \cdot \uparrow \not \in$

Y. Lee R, Childress J (1991) Assimilation of inorganic nitrogen by chemoautotrophic and methanotrophic symbioses. Appl. Environ. Microbiol. 7: 1407-1404.

١	Magee E, Richardson C, Hughes R, Cummings J ((\cdots)) Contribution of dietary protein to
۲	sulfide production in the large intestine: an <i>in vitro</i> and a controlled feeding study in humans.
٣	Am. J. Clin. Nutr. YY: 1244_1292.
٤	Marquez C, Labbate M, Ingold A, Chowdhury P, Ramírez M, Centro'n D, Borthagaray G,
٥	Stokes H ($^{\uparrow} \cdot \cdot ^{\wedge}$) Recovery of a functional class $^{\uparrow}$ integron from an <i>Escherichia coli</i> strain
٦	mediating a urinary tract infection. Antimicrob. Agents Chemother. \circ ^{1} : $10^{-10^{2}}$.
v A	Mazel D, $(7 \cdot \cdot 7)$ Integrons: agents of bacterial evolution. Nat. Rev. Microbiol. $\xi: 7 \cdot \Lambda_{-}77 \cdot .$
٩	Mazel D, Dychinco B, Webb V, Davies J (۱۹۹۸) A distinctive class of integron in the Vibrio
۱.	cholerae genome. Science ۲۸۰: ۲۰۰–۲۰۸.
۱۱	Messier N, Roy B ((\cdots)) Integron integrases possess a unique additional domain necessary for
۱۲	activity. J. Bacteriol. ١٨٣: ٦٦٩٩_٦٧٠٦.
۱۳	Minic Z, Herve G $({}^{\tau} \cdot \cdot {}^{\xi})$ Biochemical and enzymological aspects of the symbiosis between the
١٤	deep-sea tubeworm Riftia pachyptila and its bacterial endosymbiont. Eur. J. Biochem. YV1:
10	۳.۹۳_۳۱.۲
١٦	Nield B, Holmes A, Gillings M, Recchia G, Mabbutt B, Nevalainen K, Stokes H (⁽ ···)
١٧	Recovery of new integron classes from environmental DNA. FEMS Microbiol. Lett. 190:09-70.
١٨	Nandi S, Maurer J, Hofacre C, Summers A ((\cdot, \cdot)) Gram positive bacteria are a major reservoir
۱۹	of class) antibiotic resistance integrons in poultry litter. Proc. Natl. Acad. Sci. USA) ·): Y)) A-
۲.	VITT.
۲۱	Ochman H, Lawrence J, Groisman EA (⁽ ···) Lateral gene transfer and the nature of bacterial
۲۲	innovation. Nature $\xi \cdot \circ$: Y99_T · ξ .

- ۲۳ Partridge SR, Recchia GD, Scaramuzzi C, Collis CM, Stokes HW, Hall RM (۲۰۰۰) Definition
- $1 \leq 0$ of the attl¹ site of class ¹ integrons. Microbiology $1 \leq 1 \leq 1 \leq 0$.

)	
۲	Pruski A, Dixon D ((\cdots)) Toxic vents and DNA damage: first evidence from a naturally
٣	contaminated deep-sea environment. Aquat Toxicol 75: 1-17.
٤	Rodríguez-Minguela C, Apajalahti J, Chai B, Cole J, Tiedje J (۲۰۰۹) Worldwide Prevalence of
0	Class Y Integrases outside the Clinical Setting Is Associated with Human Impact. Appl. Environ.
٦	Microbiol. $\forall \circ: \circ \land \cdot \cdot = \circ \land \land \cdot$
٧	Rowe-Magnus D, Guerot A, Ploncard P, Dychinco B, Davies J, Mazel D (⁽ ···)) The
٨	evolutionary history of chromosomal super-integrons provides an ancestry for multiresistant
٩	integrons. Proc. Natl. Acad. Sci. USA ٩٨: ٦٥٢–٦٥٧.
۱.	Shoemaker N, Vlamakis H, Hayes K, Salyers A (⁽ ··)) Evidence for extensive resistance gene
11	transfer among Bacteroides spp. and among Bacteroides and other genera in the human colon.
۱۲	Appl. Environ. Microbiol. ^{TV} : °T)_°TA.
۱۳	Smalla K, Krogerrecklenfort E, Heuer H, Dejonghe W, Top E, Osborn M, Niewint J, Tebbe C,
١٤	Barr M, Bailey M, Gretaed A, Thomas C, Turner S, Young P, Nikolakopoulou D, Karagouni A,
10	Wolters A, van Elsas J, Dronen K, Sandaa R, Borin S, Brabhu J, Grohmann E, Sobecky P
١٦	$($ ^{$($} $\cdots)$ PCR-based detection of mobile genetic elements in total community DNA. Microbiology
١٧	127: 1207-1204
١٨	Stokes H, Hall R (1919) A novel family of potentially mobile DNA elements encoding site-
۱۹	specific gene-integration functions: integrons. Mol. Microbiol. ": ١٦٦٩–١٦٨٣.
۲.	Stokes H, O'Gorman D, Recchia G, Parsekhian M, Hall R (١٩٩٧). Structure and function of ٥٩-

base element recombination sites associated with mobile gene cassettes. Mol. Microbiol. $11 \quad 11: VT \quad 1-V \le 0$.

١	Stokes H, Holmes A, Nield B, Holley M, Nevalainen K, Mabbutt B, Gillings M (^(, ,)) Gene
۲	cassette PCR: sequence-independent recovery of entire genes from environmental DNA. Appl.
٣	Environ. Microbiol. ٦٧: ٥٢٤٠_٥٢٤٦.
٤	Wright M, Baker-Austin C, Lindell A, Stepanauskas R, Stokes H, McArthur J (۲۰۰۸) Influence
٥	of industrial contamination on mobile genetic elements: class \ integron abundance and gene
٦	cassette structure in aquatic bacterial communities. ISME Journal Y: ٤١٧-٤٢٨.
٧	Xu H, Davies J, Miao V ($^{(\cdot, \cdot)}$) Molecular characterization of class $^{\circ}$ integrons from <i>Delftia</i> spp.
٨	J. Bacteriol. VY: TYVI_TYAT.
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